ANALYSES AND TAXONOMIC UTILITY OF THE CEDARWOOD OILS OF THE SERRATE LEAF JUNIPERS OF THE WESTERN HEMISPHERE

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ABSTRACT

Analyses of the wood oils of the serrate leaf margined *Juniperus* of the western hemisphere (21 species, 3 varieties and 1 form) are presented. All taxa have considerable amounts of cedrol, widdrol, cis-thujopsene, α -cedrene and β -cedrene. In general, there was little correlation between cedarwood oil compositions and phylogeny in this section of *Juniperus*. *Phytologia* 91(1):117-139 (April, 2009).

KEY WORDS: *Juniperus*, Cupressaceae, wood oils, taxonomy, serrate leaf, cedrol, widdrol, cis-thujopsene, α -cedrene and β -cedrene.

The serrate leaf margined junipers of the western hemisphere appear to represent a natural sub-group of Juniperus (Adams, 2008). A phylogenetic tree (Fig. 1) shows the relationships among these junipers based on nrDNA and trnC-trnD sequences. These junipers evolved at the margins of deserts in the southwestern US and Mexico. southwestern US - Mexico, the northern-Mediterranean, and central-Asia - western China are the three centers of biodiversity of Juniperus. The serrate leaf junipers appear to be the most recent species of the genus (Adams, 2008). The group is composed of: J. angosturana R. P. Adams, J. arizonica (R. P. Adams) R. P. Adams, J. ashei Buchholz, J. a. var. ovata R. P. Adams, J. californica Carriere, J. coahuilensis (Martinez) Gaussen ex R. P. Adams, J. comitana Martinez, J. compacta (Mart.) R. P. Adams, J. deppeana Steudel var. deppeana, J. d. forma elongata R. P. Adams, J. d. forma sperryi (Correll) R. P. Adams, J. d. forma zacatacensis (Mart.) R. P. Adams, J. d. var. gamboana (Mart.) R. P. Adams, J. d. var. patoniana (Martinez) Zanoni, J. d. var. robusta Martinez, J. durangensis Martinez, J. flaccida Schlecht., J. grandis R. P. Adams, *J. jaliscana* Martinez, *J. martinezii* Perez de la Rosa, *J. monosperma* (Engelm.) Sarg., *J. monticola* Martinez forma *monticola*, *J. m.* forma *orizabensis* Martinez, *J. occidentalis* Hook., *J. osteosperma* (Torr.) Little, *J. pinchotii* Sudworth, *J. poblana* (Martinez) R. P. Adams, *J. saltillensis* M. T. Hall, and *J. standleyi* Steyermark.

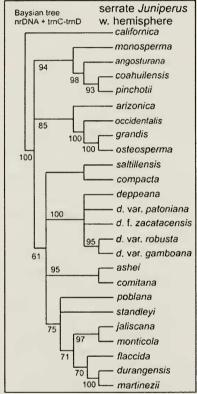


Figure 1. Phylogenetic tree of the for almost all serrate junipers (from Adams, 2008). commercially

Although the leaf essential oils of Juniperus have extensively been utilized taxonomic purposes (Adams. 1991a; Adams, 2008), the wood oils have not received much taxonomic attention. Adams (1987, Adams, 1991b) examined the wood oils of junipers from the United States as potential sources of cedarwood oil and reviewed the literature on early analyses of Juniperus wood oils.

Commercial cedarwood oils have been obtained from 3 ofCupressaceae: Juniperus (Texas, Virginia and African oils); Cupressus (China) and Cedrus (Morocco, according to Bauer and Garbe However. Texas (Juniperus ashei Buch.), Virginia (J. virginiana L.) and Chinese (putatively, Cupressus funebris Endl.) cedarwood oils account for almost all the cedarwood oil produced today 2003). (Lawrence,

heartwood oils of the Cupressaceae are well known for having the same components across the family (i.e., evolutionally conserved), so the occurrence of similar oils in different genera is not surprising. It is probably due to the conservation of the principal commercially important components (cedrol, widdrol, cis-thujopsene, α -cedrene and

β-cedrene) that the wood oil compositions have not been utilized for taxonomic purposes.

A second reason that wood oils have not been widely utilized is the difficultly in taking samples. Leaf sampling does not harm a tree. But to obtain a wood sample requires cutting down the tree, cutting off a limb, or taking a coring sample. Taking tree cores is the least destructive, but presents a problem if steam distillation is utilized to obtain the wood oil, as a core sample consists of only a few grams of wood and the oil can easily be lost on the walls of the steam distillation apparatus. Comparison of steam distillation versus solvent extraction using wood from the same tree (*J. ashei*) is shown in Table 1. Notice that the highest yield was obtained from steam distilled wood shavings and that 24 h of pentane extraction of wood chips removed only about one-half of the oil obtained by steam distillation of wood shavings. Using finely ground wood, resulted in about the same yield as using wood chips.

Table 1. Comparison using *J. ashei*_wood for yields (oven dry wt basis) and the concentrations of key components of oils obtained by steam distillation (24 h), vs. various wood chip sizes and extractions with

pentane.				
	steam	Per	tane extraction	S
	distilled	wood chips	wood from ²	ground wood
variable	shavings ¹	ext'd 24 h ²	re-ext'd +72h	$\frac{3}{24}$ ext'd $\frac{24}{1}$
per cent yield	3.7%	1.6%	1.6%	1.6%
α-cedrene	7.0	2.4	4.2	1.8
β-cedrene	1.7	1.1	2.0	1.0
cis-thujopsene	14.6	5.6	10.8	27.7
cedrol	48.1	64.3	66.1	47.2
cis-thujopsenic	acid 0.6	10.4	4.8	1.1

shavings obtained from original wood, then steam distilled 24 h.

²wood cut into 25 mm x 3 mm x 3mm wood chips, pentane extracted by shaking, 24 h. (# 9696)

³wood from 1st 24 h extraction (²), then the extracted wood chips were ground in a coffee mill and a second, 72h, pentane (# 9697) performed. ⁴original sample wood ground in coffee mill, then pentane extracted pentane for 24 h (#9700).

None of the pentane extractions gave exactly the same results as the steam distillation (Table 1). However, steam distillation can result in decomposition (Adams, 1991b), whereas solvent extraction is a very gentle method. This is shown in the marked increase of cis-thujopsenic acid in the solvent extracts (Table 1). Free acids may be dissolved in the steam condensate and return to the boiling chamber or they may decompose during distillation (Adams, 1991b). If all the wood samples are extracted in the same manner (ex. 24 h, shaking in pentane, cut to uniform sizes), solvent extraction should produce a reasonable snapshot of the profile of the wood oils.

The purpose of this paper was to present analyses of the wood oils of all the serrate junipers of the western hemisphere and evaluate these data for use as taxonomic characters.

MATERIAL AND METHODS

Samples used in the study: J. angosturana, 10.5 km e of Villa Juarez (road from Cerritos to Rio Verde), thence s 1.3 km, San Luis Potosi, Mexico, Lab # 9743 Adams 8714, J. arizonica, Rock Hound St. Park, Luna Co., NM, Lab # 9725, Adams 7637, J. ashei, 1.6 km e of Llano R., on 110, east of Junction, Kimble Co., TX, Lab #9721., Adams 5010, J. californica, 13 km n of 140 on road to Kelso, San Bernardino Co., CA, Lab # 9750, Adams 5071, J. coahuilensis, 32 km n of Alpine, TX. Jeff Davis Co., Lab # 9723, Adams 4994, J. comitana, 14 km s of Comitan and thence 14 km e on rd to Montebello, Chiapas, MX, Lab # 9737, Adams 6862, J. compacta, near the summit of Cerro Potosi, Nuevo Leon, MX, Lab # 9742, Adams 6898, J. deppeana var. deppeana, 32 km nw of Ft. Davis on Tex 118, Jeff Davis Co., Lab # 9744, Adams 4983, J. d. var. gamboana, 17 km n of Comitan on Mex. 190, Chiapas, MX, Lab # 9735, Adams 6864, J. d. var. patoniana, km 152 on Mex. 40, 52 km w of El Salto, Durango, MX, Lab # 9738, Adams 6838, J. d. var. robusta, west of Creel, Chihuahua, MX, Lab # 9728, Adams 6826, J. d. forma zacatacensis, 18 km w of Sombrerete, between km 178 & 179 on Mex. 45, Zacatecas, MX, Lab # 9740, Adams 6840, J. durangensis, nw side of Mex. 40, km 152, 52 km w of El Salto, Durango, MX, Lab # 9749, Adams 6832, J. flaccida, 20-25 km e of San Roberto Jct., on Mex. 60, Nuevo Leon, MX, Lab # 9745

Adams 6892, J. grandis, at Sonora Bridge Campground Rd., 2 km w of Jct of CA 108 and US 285 on CA 108, Mono Co., CA, Lab # 9734, Adams 5061, J. jaliscana, 19 km e of Mex. 200, on road to Cuale, Jalisco, MX, Lab # 9739, Adams 6846, J. martinezii, on La Ouebrada Ranch, 40 km n of Lago de Moreno off Mex. 85 to Amarillo, thence 10 km e on dirt rd to La Quebrada Ranch, Jalisco, MX, Lab # 9727, Adams 8709, J. monosperma, 1.6 km w of Santa Rosa, on 140, Guadalupe Co., NM, Lab # 9748, Adams 5028, J. monticola forma monticola, 1 km n of jet of Mex. 105 and El Chico Natl. Park, on road to El Chico Natl. Park (8 km ne of Pachuca), Hidalgo, MX, Lab # 9747, Adams 6874, J. occidentalis, 58 km w of Juntura on US 20, Malheur Co., OR, Lab # 9724, Adams 5085, J. osteosperma, 25 km e of Monticello, on US 666, San Juan Co., UT, Lab # 9741, Adams 5053, J. pinchotii Sudw., 10 km w of Sheffield, on I10, Pecos Co., TX, Lab # 9722, Adams 5004, J. poblana, at KM 62 on Mex. 190, 62 km s of Oaxaca, Oaxaca, MX, Lab # 9729 Adams 6871, J. saltillensis, 14 km e of San Roberto Jct., on Mex. 60, Nuevo Leon, MX, Lab # 9726, Adams 6887 and J. standlevi. 24 km nw of Huehuetango on road to San Juan Ixcoy (s of El Oro), Guatemala, Lab # 9746, Adams 6852. Vouchers are in the herbarium, Baylor University (BAYLU).

Wood samples were radially cut in 1 cm segments using a band saw. The radial sections were then cut linearly into 2 x 5 mm (x 1 cm) pieces. The wood (25 g) was placed in a 125 mL screw cap bottle to which 50 mL of pentane was added. The bottles were shaken for 24 h on a rotary shaker. The pentane extract was filtered and the pentane evaporated by use of nitrogen. The extracted wood was oven dried 48 h, 100 °C for use in the oven dry weight calculations. Percent yields were determined on an oven dry weight basis as: 100 x oil wt./(oil wt. + oven dry wood wt.). All oil samples (including commercial cedarwood oils) were dissolved in diethyl ether (10% oil solution) and stored at - 20°C until analyzed.

The extracts were analyzed on a HP5971 MSD mass spectrometer operated in the EI mode, scan time 1sec., acquisition mass range: 41-500, directly coupled to a HP 5890 gas chromatograph, using a J & W DB-5, 0.26 mm x 30 m, 0.25 micron coating thickness, fused silica capillary column, 0.2 μ L injected of a 10 % solution in

diethyl ether, and split 1/10, injector: 220 °C, transfer and MSD: 240 °C, column temperature linearly programmed: 60° - 246 °C/3 °C min. Identifications were made by library searches of our volatile oil library (10), using the HP Chemstation library search routines, coupled with retention time data of authentic reference compounds. Quantitation was by use of the HP Chemstation software. Normally one would report the data as FID values, but considering the difficultly of the peaks overlapping such that quantitation involved using single ion chromatograms to estimated the concentrations (eg. cedrol/ widdrol, etc.), it was not practical to quantitate the components by GC-FID.

RESULTS AND DISCUSSION

Tables 2 and 3 show the complete analyses of the cedarwood oils of all 25 taxa of serrate leaf margined junipers of the western hemisphere. Due to the use of a liquid extraction, considerable amounts of cis-thujopsenic acid (and other acids) were present in the extracts. It appears much of the cis-thujopsenic acid is degraded or left in the water condensate during steam distillation (Table 1). Although most taxa have considerable amounts of cedrol, widdrol, cisthujopsene, α -cedrene and β -cedrene, there are many un-identified sesquiterpenoids. However, it is surprising to find such a large number of un-identified compounds. Often, an un-identified compound was present in only one or twice taxa. It appears that there may be considerable synthesis of non-specific products in the wood. Keeling and Bohlmann (2006) discuss the defense nature of terpenoids and note that maintaining a diverse array of chemicals may be effective as a plant defense.

The major components useful in commercial cedarwood oils are cedrol, widdrol, cis-thujopsene, α -cedrene and β -cedrene. Table 4 shows the compositions of these five constituents in the 25 taxa of this study. Notice that even in phylogenetically similar taxa (Figure 1, *arizonica*, *occidentalis*, *grandis* and *osteosperma*), there is considerable variation in the amounts of these components. In fact, it seems that there is as much variation among these presumably closely related species as among other more distantly related species (Table 4).

The sum of cedrol, widdrol, cis-thujopsene, α-cedrene and βcedrene gives one some indication of the oils' utility as cedarwood oil. The sums range from 22% of the total oil (*J. jaliscana*, Table 4) to 70.4% (J. angosturana, Table 4). The species with low sums generally have considerable amounts of unknown compounds. yields ranged from 0.04% (J. deppeana var. patoniana) to 3.4% (J. standlevi). As a reference, the source of Texas cedarwood oil, J. ashei, had a 1.7% oil yield. The product of the sum of the key components x % yield (S x %, Table 4) is an index to the relative commercial potential of a species. This index varied from 2.0 (J. deppeana var. patoniana) up to 169.4 for J. angosturana with J. ashei having an index value of 104.9. Although J. angosturana (169.4), J. d. var. gamboana (128.6) and J. standlevi (143.1), all from Mexico, scored higher than J. ashei from the United States (mostly Texas), they are generally not found in large enough populations to sustain continued harvest of trees for cedarwood oil. Juniperus arizonica, J. californica, J. grandis, J. monosperma, J. occidentalis and J. pinchotii are weedy, widespread iunipers of the western US but these are either lacking a high concentration of the key compounds or their percent yields are low and do not appear suitable for commercial cedarwood oil production.

In conclusion, the cedarwood oils in this section of *Juniperus* do not seem to be useful to taxonomy at the specific level. It would be interesting to examine geographic variation within a species to determine if the wood oils might be useful for populational studies.

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occi = J. occidentalis, gran = J. grandis, oste = J. osteosperma, salt = J. saltillensis, comp = J. compacta. = J. monosperma, ango = J. angosturana, coah = J. coahuilensis, pinc = J. pinchotii, ariz = J. arizonica, Table 2. Composition of cedarwood oils from the serrate leaved Juniperus. callf = J. californica, mono

	gran oste salt comp	0.4 0.3 0.1 0.1	0.1		0.1 t 0.1 0.1				1 1	1.4 1.3 1.2 0.4	0.3	- t - 0.3	0.9 0.7 0.7 0.4	15.4 13.7 9.0 2.5	1 1	0.2 t 0.1 -	1 1	0.4 0.3 0.4 0.1	1 1		1	
-	occi	0.1	ı	,	٠	٠	1	1	•	6.0	1	,	8.0	6.4	ı	+	1	0.2	1	1	١	
- (ariz	0.1	•	•	0.4	1	-	•	•	1.9	•	9.0	9.0	14.5	1	0.3	1	0.7	ı	1	1	
	pinc	4.3	•	0.2	0.4	•	1	•	•	3.6	•	ı	1.6	12.0	•	•	•	9.0	1	_	1	
	coah	0.1	1	ı	•	0.2	•	•	0.1	=	•	ı	0.5	18.4	ı	0.3	•	0.8	0.1	•	1	
	calif mono ango coah pinc	0.1	•	1	0.1	0.1	•		,	4.		٠	0.7	17.5	ı	0.2	0.2	•	0.2	ı	ı	
	monc	9.0	1	1	,	•		1	1	0.7	1	1	0.5	0.6	1	٠	1	9.0	,		1	
	calif	0.4	1	1	1.0	1	•	•	•	1.2	1	1.3	9.0	2.2	•	٠		0.2	,	•	,	
	AI Compound	1178 naphthalene	1186 a-terpineol	1232 thymol methyl ether	1387 α-duprezianene	1390 7-epi-sesquithujene	1400 \(\beta\)-longipinene	1402 α-funcbrene	1405 sesquithujene	1410 α-cedrene	1411 2-epi-β-funcbrene	1413 \(\textit{\beta-duprezianene}\)	1419 β-cedrene	1429 cis-thujopsene	1436 isobazzanene	1449 α-himachalene	1454 (E)-β-farnesene	1464 thujopsadiene	1464 a-acoradiene	1469 \(\beta\)-acoradienc	1470 10-epi-β-acoradiene, isomer	

1	1	1	0.1	•	1	•	•	1	•	0.8	1	1	•	1	•	1	•	•	•	1	1	0.2	•	4.1
٠	٠	٠	ı	١	٠	0.3	٠	•	1.7	1.2	0.1	1	+	,	٠	0.4	1	٠	•	,	1	•	ı	1.0
	0.2		•	,		0.2	0.2	•		Ξ	-	+	+		ı				•	-		,	•	1.0
ı	0.3		0.2	1	0.1	0.1	0.2		0.1	4.	0.2	+	٠	-	٠	-		r	٠.	0.1		•	٠	1.2
0.1	,	1	0.1	ı	0.2	ı	1		-	6.0			~	0.2	,	,				0.3			0.1	6.0
ı	0.5			ı	0.3	0.5	0.4		-	1.7	0.5		0.3	,	٠	•		1.3		ı		•		1.0
		0.5	,			,	6.0	•		2.4		1.3	0.7						0.5			•	0.4	1.4
0.1				0.1	ı	ı	0.3			1.2	0.2		,			0.3	9.0				9.0	1	0.4	2.0
1	0.2	•		,	1		0.7			0.3	0.1		ı	t	0.3	,	1			0.1		ı	•	6.0
ı								0.2	,	1.3			0.5			0.5			8.0	ı	0.2		0.3	1.7
ı	,		9.0		ı		0.4	ı		4.8	1					0.3			0.3	ı	1		•	2.2
1479 ar-curcumenene	1481 γ-himachalene	1485 11-αH-himachala-1,4-diene	1489 β-selinene	1498 β-alaskene	1498 α-selinene	1499 pseudowiddrene	1500 β-himachalene	1502 trans-β-guaiene	1503 α-chamigrene	1504 cuparene	1512 α-alaskene	1514 butylated hydroxy toluene*	1521 trans-calamenene	1522 8-cadinene	1532 γ-cuprenene	1536 italicene ether	1540 sesquiterpenol <u>43</u> ,44,95,220	1541 sesquiterpenol 43,97,205,220	1541 8,14-cedranoxide	1542 \delta-cuprenene	1546 elemol	1562 longicamphenylone	1582 caryophyllene oxide	1589 allo-cedrol
	ar-curcumenene 0.1	ar-curcumenene 0.1 0.1 γ-himachalene 0.2 0.5 0.5	ar-curcumenene 0.1 0.1 - γ -himachalene 0.2 0.5 - 11- α H-himachala-1,4-diene 0.5	ar-curcumenene 0.1 0.1 - γ -himachalene 0.2 0.5 - γ -himachala-1,4-diene 0.5 γ -selinene 0.6 0.1	ar-curcumenene 0.1 0.1 - γ -himachalene 0.2 0.5 - γ -himachala-1,4-diene 0.5 γ -b-selinene 0.6 0.1 0.1 0.1	ar-curcumenene 0.1 0.1 γ-himachalene 0.2 0.5 11-αH-himachala-1,4-diene 0.5 0.5 6β-selinene 0.6 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1 0.1	ar-curcumenene 0.1 0.1	ar-curcumenene 0.1 0.1 0.1 0.1 0.1 0.1					

comp			54.2		1.3		0.7	0.4	0.5	ı		1	1		,		ı	1.9	t	1			,	,	
salt	0.2	+	46.9	0.3	0.3	٠	8.9	1.3	,	•	0.5	١	1	ı	6.0		١	٠	١	١	ı	١	1	1	1
oste	0.3	-	36.3	•	0.3	•	3.1	8.0	ı	1	•	•	3.7	1.5		٠	١	٠	1	٠	1.9	١	٠	1	•
gran	0.2	٠	41.8	0.4	Ξ:	•	3.1	0.8	•	0.3	6.0	'	٠	•	•	١	2.3	•	•	ı	'	٠	'	٠	1
occi	0.3	+	43.3		1.2	1	4.0	1.0	•	r	•	0.0	0.4	•	0.7	1	•	2.0	•	1	•	ı	1.4	٠	٠
ariz	0.3	7.6	3.1	•	-	•	3.4	6.0	1	1	,	•	٠	t	•	1	٠	2.2	•	1	'	•	4.4	٠	•
pinc	0.1	4.2	38.6	-	0.4	٠	3.2	1.2	•	•	•	1.9	0.2	-	٠	•	•	٠	9.0	1.9	ł	•	,	•	•
coah	٠	1	17.4	0.5	9.0	9.0	5.9	9.1	•	•	1.7	٠	3.2	1.4	٠	١	١	•	٠	1.6	1	•	•	0.7	1
mono ango coah pinc	•	-	51.0	0.5	9.0		3.6	8.0	•	,	•	8.0	9.0		٠	8.0	1	ı	•	1		0.7	,	•	1
mono	+	7.1	14.1	1.3			5.7	1.8		ı	•	1.5	6.9	1.4	•	,	•	1		3.2		•	٠	•	•
calif	0.5	11.7	11.7	9.0	,		8.0	0.7		8.0		•	1.8	1	1.3	1	4.1		,	1	ı	•	•	,	0.3
AI Compound	1595 cis-dihydro-mayurone	1599 widdrol	1600 cedrol	l612 β-biotol	1618 epi-cedrol	1622 sesquiterpenol <u>111</u> ,43,69,220	634 α-acorenol	636 β-acorenol	1639 1,7-di-epi-α-cedrenal	1641 3-iso-thujopsanone	648 sesquiterpenol 119,43,204,220	1649 sesquiterpenol, <u>43</u> ,119,71,220	(649 β-eudesmol	652 α-eudesmol	1658 selin-11-en-4α-ol	1660 sesquiterpen-ol 135,79,220	1662 sesquiterpenal 43, 123, 236	1663 sesquiterpenal <u>135,</u> 91,69,236	1665 intermedeol	1665 sesquiterpenal 43,123,95,236	1666 sesquiterpenal <u>43</u> ,69,135,236	l666 sesquiterpenol 119,41,93,220	1667 sesquiterpenol 105,91,119,218	1668 sesquiterpenol <u>41</u> ,69,91,220	1675 cadalene

comp	,		0.4	8.0			1.0		1.2			0.4			,	0.4			,	0.7	0.5	•	,		
salt	•	٠	ı	•	١	2.6	•	5.6	•	2.5	1	١	•	ı	•	٠	ı	•	•	8.0	•	٠	1	١	٠
gran oste salt		•	Ξ	٠	1	٠	3.5	2.5	•	1.7	1	•	٠	Ξ	•	•	•	1.0	٠	٠	0.3	•	١	1	•
		-	0.4	•	•	•	2.6	2.4	•	•	8.0	1	•	1	-	-	1	•	•	0.5	0.3	0.2	ı	١	•
occi	ı	0.1	•	•	•	•	•	1	•	1	•	1	•	1	1	1.6	•	Ξ	1	1	•	•	9.0	1	0.5
ariz		١		٠	٠	٠	3.7	•	•	٠	0.0	•	•	•	١	٠	١	•	•	1.2	•	٠	٠	'	'
pinc	0.4	ı	3.2	•	٠	•	•	ı	3.5	•	1.5	1.2	1	1	0.5	•	1	•	0.4	٠	,	9.0	•	٠	•
coah	0.5	0.5	•	•	•	•	4.0	4.4	•	Ξ:		•	Ξ:	ı	•	2.3	1	•	1	٠	٠	٠	•	٠	4.8
mono ango coah ninc	0.2	0.2	ı	•	١	٠	3.6	4.8	1	•	٠	•	٠	٠	٠	١	٠	8.0	٠	. 0.5	٠	١	٠	9.0	٠
mone	1.5	١	1	ı	8.0	ı	1.5	4.3	4.3	1	2.3	2.7	ı	٠	2.2	ı	1.0	٠	8.0	i	٠	1.7	٠	•	•
calif	١.	1	•	1	٠	2.5		•	18.5	٠	3.2	4.2	,	٠	2.8	- 9	•		8.0	4.6	•	•	0.4		-
AI Compound	33 e	1685 a-bisabolol	1688 8-cedren-13-ol	1689 sesquiterpenol <u>43</u> ,121,132,222	1692 junicedranol	1694 sesquiterpenol, 41,91,135,220	1699 sesquiterpenol 135,79,105,220	1701 cis-thujopsenol	1703 mayurone	1708 cis-thujopsenal	1709 sesquiterpenal 123,41,218	1714 sesquiterpene 43,135,207232	1716 sesquiterpenol 43,135,207,222	1724 (Z)-nuciferol	1726 sesquiterpenal 43,137,218,236 2.8	1729 sesquiterpenal 119,147,162,236	1748 sesquiterpenal <u>123,55,41,234</u>	1751 sesquiterpenol 105,41,147,220	1751 cuparenal	1762 \(\beta\)-acoradienol	1765 \(\beta\)-costol	1766 sesquiterpenal 41,55,107,234	1773 α-costol	1789 sesquiterpenol 123,41,135,220	1793 sesquiterpenal 189,43,207,236

comp		2.8		0.5								0.1	15.2	0.7	,		•	,	9.0		,	•	,	1	
salt		١	٠	4.1	•	٠	•	1.3	٠	٠	•	5.8		٠	1	٠	<u> </u>	ı	9.1	0.7	١	١	١	ı	
oste		1.2	١	9.0	1	٠	0.5	٠	•	٠	•	9.9	2.7	0.1	•	٠	0.7	•	1	•	,	1	1.5	1	
gran	9.0	ı	,	8.0	١	•		1.3	•	٠	٠	5.5	2.1	0.3	٠	0.5	8.0	1	4.	١	٠	,	٠	8.0	
occi	9.0		•	0.1	٠	,	•	0.5		٠	٠	5.9	2.7		•	ı	0.7	٠	8.0	٠	1	٠	•		
ariz	2.8	a		8.0		٠	1	1.0			2.1	6.7	٠	٠		٠	•	٠	1.4	٠	٠	•	٠	,	
pinc	0.7	1		9.0	٠	٠			•		1	0.7	•		•		٠	٠	•	•	•		•		
coah	•	٠	•	Ξ:	,	,	8.0	٠	,			<u>8:</u>	3.1	1.0	ı	9.0	ı		,	,	8.1	ı	•	١	
ango				8.1	,		0.3	٠	0.5	1		6.0	ı					٠					ı		
mono ango coah pinc		3.1		٠	8.0		9.0	1		١	,	3.6	8.	-	•	3.0				•		4.		1	
A1 Compound calif	al 148,131,41,218 -	1797 sesquiterpenal 43,189,207,236 -	1801 sesquiterpene 95,91,171,232 1.3	1802 sesquiterpol <u>121</u> ,136,177,220	1805 sesquiterpenal 121,136,177,234 -	1810 sesquiterpenal 55,135,234 0.8	1813 cryptomeridiol	1815 sesquiter acid 149,91,105,234 1.0	1831 sesquiterpenol 91,105,135,220 -	1836 sesquiterpene <u>43,119,190</u> 0.7	1837 sesquiterpene 119,161,43,191	1863 cis-thujopsenic acid 1.7	1889 cedrane-8S,14-diol 2.2	1897 cedrane-8S,13-diol -	1911 sesquiterpenal 41,123,211,234 1.6	1917 sesquiterpenal 41,123,149,234 -	1929 sesquiterpenal 121,136,191,234 -	1932 cuparenic acid 0.6	1945 sesquiterpenal 121,136,191,234 -	1953 sesquiterpene 43,135,181,256 2.4	1959 nootkatin	1959 sesquiterpene <u>135</u> ,43,181,256 -	1960 sesquit. acid 121,136,191,234 -	1962 sesquiterp. acid 41,43,135,256 -	

o coah pinc ariz occi gran oste salt comp		•	0.4
salt	•	•	1
oste	•	ı	,
gran	,	1	0.1
occi	•	ı	1
ariz	5.3		-
pinc	,	,	1
coah	Ξ	,	•
ango		1	
calif mono ango		,	-
calif	- +	+	
AI Compound	1964 sesquiterp. acid 135,43,163,23	2158 octadecanoic acid	2331 trans-ferruginol

considered a non-natural product, RI = Arithmetic Index on DB-5 (= SE54). column. Compositional values less than For unknown cpds., base ion (100%) is underlined, next ions are major ions, *butylated hydroxy toluene is normally 0.1% are denoted as traces (t). Unidentified components less than 0.5% are not reported.

Table 3. Composition of cedarwood oils from serrate leaves Juniperus. depp = J. deppeana, dpat = J. d. var. patoniana, $pobl = J.\ poblana,\ stan = J.\ standleyi,\ jali = J.\ jaliscana,\ mont = J.\ monticola,\ flac = J.\ flaccida,\ dur = J.\ durangensis,$ mart = J. martinezii.

AT Commond	denn	denn dnat dzac		drob	drop dgam ashe comt pobl	ashe	comt		stan	jali	mont flac		dur	mart
Collipound											,	,	•	0.1
932 α-pinene								,						
969 sabinene			ı	1		ı			ı		ı	ı		
1024 limonene		ı		ı	ı		,		t	ı				0.1
	0.4	4.	9.0	0.3	0.1	0.1	0.1	0.1	0.4	0.1	0.5	0.2	0.3	0.1
1298 carvacrol		0.1	ı			•					ı			
1380 2-eni-α-funebrene	0.1	•	0.1	,				ı	0.1		ı	•		ı
	0.3	0.1	0.4	0.2	0.1	1	-	0.1		,	0.5	0.1	0.2	0.2
		٠		٠	1		0.4	•	ı	,				
1390 7-epi-sesquithujene	•	•		•	ı			٠	0.1		ı		ı	
	•		0.1	0.1	+	ı		,		•	,	•		
- ~				-	1	-			•		ı			•
	,	,	,	ı		,	,		0.1	1	ı	,	ı	ı
	7.6	1.5	4.1	3.9	2.5	9.0	0.5	3.3	5.6	5.6	1.9	3.5	1.2	2.0
	,	+	•	+		ı		ı		9.0				,
1417 (E)-caryophyllene	•		•	•	ı	ı	0.1		ı			1	•	. (
	2.0	8.0	1.4	6.0	1.0	0.4	0.2	6.0	1.2	0.7	0.0	1.0	6.0	6.0
		•	ı	٠	•	,	0.5	,	,	,	ı	ı	ı	
	9.1	7.2	10.3	6.3	5.5	12.0	5.9	9.9	10.3	9.7	14.3	8.8	7.9	8.9
	,	٠	+	1	,			1	0.1					

ne 0.2 ne 0.3 nala-1,4-diene 0.2 ne 0.6 e 0.6 e 0.6 e 0.6 e 0.6 e 0.7	Compound	depp dpat	dpat	dzac	drob	dzac drob dgam ashe	ashe	comt pobl	pobl	stan	jali	mont flac	flac	dur	mart
e + +	chalene	0.2		-	0.1		0.1		,		0.1		0.1		1
e 0.6 0.1 0.3 0.2 0.1 - 0.2 0.2 0.1 0.1 0.2 0.3 0.3 0.3 0.3 0.3 0.3 0.3 0.3 0.3 0.3	arnesene			-		,	,	ı							ī
1.1 - 0.6 0.6 t - 0.7 0.3 0.7 0.1 0.1 0.3 0.3 0.7 0.1 0.3 0.3 0.7 0.4 0.2 t - 0.1 0.1 0.1 - 0.4 0.2 0.3 0.7 0.4 0.2 - 0.3 0.7 0.4 0.2 - 0.3 0.7 0.1 0.1 0.1 0.1 0.3 0.3 0.7 0.4 0.2 - 0.3 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1	sadiene		9.0	0.1	ı		0.3	0.2	0.1		0.2	0.3	0.1	0.2	0.3
0.3 - 0.5 0.2 t - 0. 0.1 0.1 - - 0.3 0.7 0.4 0.2 - 0 - 0.1 0.1 - 0.3 - 1 t - 0.2 - 1 t - 0.1 0.1 - 1 ala-1,4-diene	adiene	-:	ı	9.0	9.0	-			0.3	0.7	0.1	0.3	0.1	0.2	
e - 0.3 0.7 0.4 0.2 - 0.1 0.1 0.4 0.4 0.3 0.3 0.3 0.3 0.2 0.4 1 0.3 0.1 0.1 0.1 0.4 0.4 0.3 0.3 0.3 0.3 0.3 0.3 0.3 0.3 0.3 0.3	adiene	0.3		0.5	0.2	-			0.1	0.1	1	t	-		0.1
e 0.2 - t - 0.1 0.1 0.3 0.3	nigrene	1	0.3	0.7	0.4	0.2			0.1	,	0.4	0.3	-	-	0.3
0.3 - t - 0.3 - 1 - 1 - 0.3 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 1 - 0.5 - 0.	cumenene				0.2		+	1	0.1	٠		ı	+	٠	0.1
achala-1,4-diene	achalene	0.3		-	,	,	0.3		,	,	٠	,	ı		ı
trene 0.6 0.1 0.5 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1 - 1		ı	ı	ı				-							
10.2 0.1 0.4 1 1 1 1	nene	ı	ı	•				0.5				,			,
trene 0.6 - 0.7 0.2 - 0.6 - 0.6 - 0.7 0.2 - 0.2 - 0.6 - 0.7 - 0.2 - 0.2 - 0.6 - 0.4 - 0.5 0.2 0.2 - 0.4 - 0.4 - 0.5 0.2 0.2 - 0.4 0.4 0.5 0.2 0.2 - 0.4 0.5 0.2 0.2 - 0.4 0.5 0.3 0.1 0.2 0.3 0.7 - 0.4 0.5 0.1 0.1 - 0.2 0.3 0.7 - 0.4 0.5 0.1 0.1 - 0.2 0.2 0.3 0.7 - 0.4 0.5 0.1 0.1 - 0.2 0.2 0.3 0.1 0.1 0.1 0.6 0.1 0.1 0.3 - 0.5 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1	kene	0.2			0.1	ı	1		+	•					
threne 0.6 - 0.7 0.2 - 0.6 - 0.6 - 0.6 one	iene	ı	1	0.7				0.4					٠		,
ene	widdrene	9.0	1	0.7	٠		0.2		9.0		0.7	0.5	ı	1	0.4
t - t t - t - t - t - t 0.2 (1.6 1.3 2.1 1.2 0.8 1.2 0.3 1.1 0.6 1 0.7 - 0.4 0.5 0.1 0.1 0.2 (ydroxy toluene* - 0.7 0.2 1 inene	achalene	0.5	0.4		1.2	0.5	0.2	0.2	1	0.4	9.0		9.0	0.4	0.5
1.6 1.3 2.1 1.2 0.8 1.2 0.3 1.1 0.6 10.7 - 0.4 0.5 0.1 0.1 - 0.2 0.2 0.2 0.2 0.3 0.1 0.1 - 0.2 0.2 0.2 0.3 0.1 0.1 - 0.2 0.2 0.2 0.3 0.1 0.1 0.1 0.2 0.2 0.2 0.3 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1	nigrene	+		-	~		-			0.2	0.2				-
0.7 - 0.4 0.5 0.1 0.1 - 0.2 or over the contract of the contra	ne	9.1	1.3	2.1	1.2	8.0	1.2	0.3	=	9.0	1.0	0.1	1.0	3.0	8.0
ydroxy toluene* - 0.7 t inene 0.1 0.3 - 0.5 0.1 0.1 0.1 e t e	kene	0.7		0.4	0.5	0.1	0.1			0.2	0.3	0.2	0.2	,	0.2
/ toluene* - 0.7 t t 0.1 0.3 - 0.5 0.1 0.1 0 0.1 0.1 0 0.1 0.1 0	nene				ı	ı		0.2				,			
0.1 0.3 - 0.5 0.1 0.1 0 0.5 0.1 0.1 0 0.1 0.1 0	ted hydroxy toluene*	,	0.7		٠				٠	,	,		•		
0.1 0.3 - 0.5 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1 0.1	α-selinene	•	,				-								
0.1	calamenene	0.1	0.3	•	0.5	0.1	0.1	0.1	0.1		٠	,	0.1		
0.1	nene	1	ı		1	,	0.1	0.1				ı	,		0.1
	enene	0.1		,						0.1	0.1				0.3
italicene ether - 0.5 0.1	the ether	ı	0.5		1	1						0.1	•	0.7	,

AI Compound	depp dpat	dpat	dzac	drob	dgam ashe		comt pobl		stan	iali	mont flac		dur	mart
=		8.0	1:1	,		٠			٠				ı	
		•	•	•	0.7					,		,		1
	0.4	ı	,				,		,			•	. :	
			٠	٠	,			0.1		0.7	0.1		0.5	
		•	,			0.1		,				ı		0.2
	ı		•	,		•	0.2						ı	
	1	ı	,	,	,	0.1	,		ı	ı	,	1	,	,
	0.3	1.0	0.3	0.3	0.5	4:1	1.2	0.3	1.0	0.3	1.0	0.4	2.0	9.0
	0.1	0.3	0.5	0.2	0.2	-	٠	0.3	0.2	0.5	0.4	0.3	0.4	,
	12.0	10.0	16.0	15.6	0.1	12.0	•	20.6	25.2	10.0	7.0	32.0	5.0	3.7
	25.7	30.4	1.3	2.0	55.2	36.7	43.2	21.0	2.8	Ξ	10.4	15.9	21.9	34.0
	0.4	0.5		0.2	•		6.0	٠				0.5	•	,
	,	1	•	0.2	٠	,	•	0.3	ı	0.5	ı	ı	ı	,
	1.2	0.7	•	0.5	0.4	0.5	,	9.0	8.0	0.2	1.2	1.4	8.0	1:1
	,	٠	٠	٠	٠	0.5	,	ı		•		ı		
	ı	•	,	1	١	1	0.5	•	1	ı	ı		1	,
	3.7	3.4	1.2	3.3	1.6	4.9	1.2	2.1	19.7	8.0	4.3	1.3	3.6	2.3
	1.6	-:	6.0	1.4	0.7	2.0	0.4	1.0	3.2	0.4	1.6	1.0	-:	0.1
	•	•	٠	٠	•	•	•	١		٠	ı	1.6	•	. !
	١	ı	•	٠	1	,	0.5	0.5	•	0.5	1	0.4	•	0.7
	٠	•	٠	١	٠	•		٠	٠	,	•	٠	0.8	1
	-	١	ı	٠	0.3	2.7	•	0.5	•	1	•			
	,	9.0	٠	,	٠	٠	1	•	•	•				1
	1	9.0	1	1	1	1	•	•	1	1	•	1	1	

Compound depp dpat
ı
,
0.3
ı
7.(
,
0.2
,
ı
0.1

mart		,	4.1		2.8	1	•	0.5	,	,	•	•	•	1.2	•	1	1	,	ı	1	6.0	1	,	1
dur	0:1	٠	3.0	•	٠	9.7	•	-:	4.1	1		•	0.5	•	9.0	٠	ı	١	١	1	1	٠	3.9	•
flac	,	•	5.7		2.3	•		9.1	1	4	1	٠	1	2.0	,	٠	1.2	٠	١	1	١	1	٠	1
mont flac		٠	4.1			3.8	•	1.4				1.3	•	1	•	•	•	1	1	•	6.0	,	0.3	
jali		•	5.4			,	2.3	0.5	0.3	•	•	,		•	,		0.7	ı	1	•	ı	١		0.5
stan	,	2.9	,	٠	4.0	٠		,	,	,	٠	•	٠	1		•	•	1	2.4	ı	•	1		,
pobl	,		5.5	,	5.6	1		١	٠	1.2		0.5	٠			,	1	•	٠	1	1.3	•	•	•
comt pobl	ı		Ξ	8.9	1.4	1		ı					ı	•		1.0	ı	1	ı	ı	•	2.9		
		,	8.0		4.4		2.2	,		1	٠	1	,		·	,					6.0	9.0		
dgam ashe		,	8.8	,	2.5		,	2.1				1		ı		,	1.5		,	,	٠	,		
drob	,	,	4.5		3.8	•		1.8	0.7	ı		,	ı				1.4	ı		ı	ı	ı	1	1.7
dzac		,	5.1	ı		4.	,	2.2	8.0			1		ı			8.0		ı	1.2		1		1.6
			5.6			5.8	,	1.2	2.2									0.7	ı	,		,	4.	
depp dpat	,	ı	1.4		,	1.5		1.3		,		,	ı				0.4			0.5	ı	ı	ı	
AI Compound	1689 sesquiterpenol 43,121,132,222	1694 sesquiterpenal 135,71,41,220	1699 sesquiterpenol 135,79,105,220	1700 sesquiteral 107,91,119,218	1701 cis-thujopsenol	1703 mayurone	1708 cis-thujopsenal	1709 sesquiterpenal 123,41,218,220	1714 sesquiterpene 43,135,207,232	1714 sesquiterpenal <u>123</u> ,55,206,236	1716 sesquiterpenol 43,135,207,222	1724 (Z)-nuciferol	1726 sesquiterpenal 43,137,218,236	1729 sesquiteral 119,147,162,236	1736 sesquiterpenal 43,123,132,218	1745 γ-costol	1751 sesquiterpenol 105,41,147,220	1751 cuparenal	1753 sesquiterpenol 105,41,79,222	1759 sesquiterpenal 119,93,238	1762 β-acoradienol	1765 β-costol	1766 sesquiterpenal 41,55,107,234	1771 sesquiterpenal <u>119</u> ,93,160,234

AI Compound	depp dpat		dzac	drob	drob dgam ashe	ashe	comt pobl	ldod	stan	jali	mont	flac	dur	mart
1773 α-costol	,	0.5	,			0.2	8.1	,	١	'	١	ı	ı	
1774 sesquiterpenal 91,41,69,235			ı	0.7		٠			٠			•	٠	
1779 14-hydroxy-α-muurolene							•	,	,	•		•		9.0
1781 sesquiterpenal 119,93,234				9.0	1	٠		,	٠	<u>«</u> .		٠	٠	
1784 sesquiterpal <u>159</u> ,119,220,234	ı					•	1	,		6.0		,	,	,
1785 sesquiterpal <u>121</u> ,136,41,218		ı	ı			9.0		1						
1787 sesquiterpal 119,93,234		٠				٠			,	9.0	٠	ı	٠	,
1793 sesquiterpenal 189,91,133,234		,	,	2.5	1.3	•		0.1	١	٠	٠	1.3	•	0.5
1793 sesquiterpenal <u>189</u> ,43,207,236		,	1.9		,	•			٠	3.9	٠	,	•	,
1793 sesquiterpenal 41,55,79,232		1.2	1		1			,	٠	,		•		,
1794 sesquiterpenal <u>91</u> ,79,105,218			,			•	1.9		1			•	•	
1796 sesquiterpenal 148,131,41,218	1	ı	ı		0.4	2.0		1.7		,		1	1	,
1797 sesquiterpenal 43,189,207,236		1	ı		,	ı	ı	١	,	ı	9.0		1.3	
1799 sesquiterpal 189,148,165,236	1		ı	2.4				,	1	٠	٠	1	•	,
1802 sesquiterpol <u>121</u> ,136,177,220		0.7	Ξ:		4.	1.9	9.0	2.1	1.9		٠	1.8	0.5	8.0
1803 sesquiterpal <u>189</u> ,121,43,236						,	,			3.9				
1805 sesquiterpenal <u>121</u> ,136,177,234	0.1		,	5.4		ı		,		٠	٠			,
1806 nootkatone	ı						1.0			•	ı	•	•	
1810 sesquiterpenal 41,91,95,234		1	•				4.1			1			ı	,
1815 sesquiter. acid 149,91,105,234	3.0	0.5	6.0								2.2			
1821 sesquiterpenal <u>123</u> ,43,205	,	9.0				•	,		•	,				
1821 sesquiacid <u>149</u> ,105,191,234			,	=	,					•		1.0	,	,
1837 sesquiterpene 119,161,43,191							,	,	,	٠	0.3			
1863 cis-thujopsenic acid	7.1	2.3	17.8	8.3	4.4	2.1	0.4	0.9	2.4	5.4	11.0	5.6	4.1	6.4
											I	١]

depp dpat
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inol	Al Compound	depp	dpat	dzac	drob	dgam a	she	comt	lgod	stan	lalı	mont	flac	dur	mari
0.2	1984 sesquitacid 135,43,181,256			1					,	•	11.6	4.7	•	•	
	2077 sesquiterpal 59,149,105,234	•		1	ı	,	,			1.0	1		•	٠	
	2082 sesquiterpene 43, 202/203,234	ı		•	0.7	1		ı	,	ı	1	,		٠	ı
0.2	2141 oleic acid	•	1	1				,		•	•		ı	ı	2.7
. 0.2	2158 octadecanoic acid					•			ı					•	2.0
	2269 sandaracopimarinol		0.2			1			+					٠	•
, , , , , , , , , , , , , , , , , , , ,	2314 trans-totarol	•	ı	,	•				ı	ı	1			0.1	
0.2 0.2 - 0.4 0.3 0.3 -	2331 trans-ferruginol	0.2	0.2	,	0.2	•		0.4	0.3	0.3	•	+	0.2	1	1

Table 4. Comparison of the major oil components among taxa. ACDR = α -cedrene, BCDR = β -cedrene, THJP = cis-thujopsene, WDRL = widdrol, CDRL = cedrol, CTJA = cis-thujopsenic acid. %yld = % yield on oven dry wt basis (oil/(extracted wood, dried 48h, 100°C).

	ACDR	BCDR	THJP	WDR	LCDRL	Sum	%yld	S x %
J. californica	1.2	0.6	2.2	11.7	11.7	25.4	0.2	5.0
J. monosperma	0.7	0.5	9.0	7.1	14.1	31.4	0.3	9.4
J. angosturana	1.4	0.7	17.5	t	51.0	70.6	2.4	169.4
J. coahuilensis	1.1	0.5	18.4	-	17.4	37.4	1.6	59.8
J. pinchotii	3.6	1.6	12.0	4.2	38.6	60.0	0.2	12.0
J. arizonica	1.9	0.6	14.5	7.6	3.1	27.7	0.6	16.6
J. occidentalis	0.9	0.8	6.4	t	43.3	51.4	0.5	25.7
J. grandis	1.4	0.9	15.4	-	41.8	59.5	0.2	11.9
J. osteosperma	1.3	0.7	13.7	t	36.3	52.0	0.3	15.6
J. saltillensis	1.2	0.7	9.0	t	46.9	57.8	0.1	5.8
J. compacta	0.4	0.4	2.5	-	54.2	57.5	1.0	57.5
J. deppeana	7.6	2.0	9.1	12.0	25.7	56.4	0.3	16.9
patoniana	1.5	0.8	7.2	10.0	30.4	49.9	0.04	2.0
zacatacensis	4.1	1.4	10.3	16.0	1.3	33.1	0.5	16.6
robusta	3.9	0.9	6.3	15.6	2.0	28.7	0.7	20.1
gamboana	2.5	1.0	5.5	0.1	55.2	64.3	2.0	128.6
J. ashei	0.6	0.4	12.0	12.0	36.7	61.7	1.7	104.9
J. comitana	0.5	0.2	2.9	-	43.2	46.8	1.0	46.8
J. poblana	3.3	0.9	6.6	20.6	21.0	52.4	1.8	94.3
J. standleyi	2.6	1.2	10.3	25.2	2.8	42.1	3.4	143.1
J. jaliscana	2.6	0.7	7.6	10.0	1.1	22.0	1.4	30.8
J. monticola	1.9	0.9	14.3	7.0	10.4	34.5	0.7	24.2
J. flaccida	3.5	1.0	4.8	32.0	15.9	57.2	0.9	51.5
J. durangensis	1.2	0.9	7.9	5.0	21.9	36.9	0.2	7.4
J. martinezii	2.0	0.9	6.8	3.7	34.0	47.4	1.2	56.9